Concanavalin A (ConA)

-AcalephFluor633

User Manual

Catalog # CRG1146

AcalephFluor633 labled Concanavalin A (ConA) can bind

alpha-D-mannosyl and alpha-D-glucosyl residues from B-Glycans

For research use only. Not for diagnostic or therapeutic procedures.

I. INTRODUCTION.....2
II. MATERIALS REQUIRED BUT NOT PROVIDED.....3
III. WORKING SOLUTION PREPARATION.....4
IV. ASSAY PROCEDURE.....5
V. FLUORESCENCE SPECTRUM.....7

I. INTRODUCTION

Concanavalin A (ConA) is a lectin that binds specifically to certain structures found in various sugars, glycoproteins and glycolipids. ConA is widely used in biology and biochemistry to characterize glycoproteins and other sugar-containing entities on the surface of various cells. ConA binds specifically alpha-D-mannosyl and alpha-D-glucosyl residues (two hexoses differing only in the alcohol on carbon 2) in terminal position of ramified structures from B-Glycans. It has 4 binding sites, corresponding to the 4 subunits.

II. MATERIALS REQUIRED BUT NOT PROVIDED

1. PBS (1X)
2. 4% Formaldehyde
3. 0.5% Triton X-100
4. DAPI
5. Antifade Mounting Medium

III. WORKING SOLUTION PREPARATION

Stock Solution (200X): Dissolve the lyophilized powder in 300 µl ddH_2O for the 300 Assays size or 50 µl ddH_2O for the 50 Assays size.

Working Solution: Dilute 1 µl fluorescent ConA stock solution in 200 µl PBS before use.

(For fluorescent ConA, the recommended dilution ratio is 1:100 - 1:200, one time experiment is equivalent to 1-2 µl stock solution in a total staining volume of 200 µl.)

Note: The dilution ratio can be adjusted appropriately according to the experimental effect.

Note:

The reconstituted conjugate solution can be stored at 2-8 °C for short-term storage or at -20 °C for long-term storage. Avoid repeated freeze-thaw cycles.

IV. ASSAY PROCEDURE

Staining fixed cells (24-well plate/Cell slide)

1. Wash cells 3 times with PBS.
2. Fix cells on ice with 4% formaldehyde solution in PBS for 15 minutes.

Note: For fixed cell membrane staining, it is recommended to stain without the permeabilization step. A permeabilization step after fixation can facilitate staining intracellular compartments such as Golgi and Endoplasmic Reticulum (ER) structures.

3. Wash cells 3 times with PBS.
4. Add 200 µL ConA Working Solution, incubate for 10-30 minutes at room temperature.
5. Wash 2-3 times with PBS.
6. Stain the nucleus (optional): Add DAPI staining solution to cover the cells on the sample, perform cell nucleus staining for 5-10 minutes. Wash the sample 2-3 times with PBS, each time for 5 minutes.
7. Image using fluorescence microscopy. Fluorescent ConA are photostable enough to image in PBS, but for best results we recommend mounting with antifade mounting medium.

Staining living cells (24-well plate)

1. Wash cells twice with PBS buffer.
2. Add 200 µL ConA Working Solution.
3. Incubate cells with ConA Working Solution for 10-30 minutes at 37°C.
4. Wash cells twice with PBS buffer.
5. Stain the nucleus (optional): Add DAPI staining solution to cover the cells on the sample, perform cell nucleus staining for 5-10 minutes. Wash the sample 2-3 times with PBS, each time for 5 minutes.
6. Image using fluorescence microscopy.

Tissue section staining

1. Pre-treatment of tissue sections

Frozen sections were left at room temperature for several minutes to restore to room temperature; the sections were immersed in the fixative at room temperature for 10-15 minutes, then removed and naturally dried, and then rinsed with pure water or PBS to remove the residual fixative on the tissue.

Paraffin sections were placed in the dewaxing solution for 10 minutes, then immersed in anhydrous ethanol for 10 minutes, 95% ethanol for 10 minutes, 80% ethanol for 10 minutes, and distilled water for rinsing.

2. Antigen retrieval, sections were placed in Tris-EDTA pH 8.0, and heated for 15 minutes for retrieval. After natural cooling, the slides were placed in PBS and washed 3 times, each time for 5 minutes.

3. Add 50-100 µl of ConA Working Solution to cover the tissue, incubate at 37°C in the dark for 30 minutes.

4. Wash the sample 3 times with PBS, each time for 5 minutes.

5. Stain the nucleus (optional): Add DAPI staining solution to cover the cells on the sample, perform cell nucleus staining for 5-10 minutes. Wash the sample 2-3 times with PBS, each time for 5 minutes.

6. After slightly drying the section, add Antifade Mounting Medium on the sample and use a suitable cover slip for mounting.

7. Image using fluorescence microscopy.

V. FLUORESCENCE SPECTRUM

Conjugation	Excitation	Emission
Concanavalin A (ConA) - AcalephFluor405	404 nm	431 nm
Concanavalin A (ConA) - AcalephFluor488	491 nm	512 nm
Concanavalin A (ConA) - AcalephFluor532	525 nm	554 nm
Concanavalin A (ConA) - AcalephFluor555	555 nm	565 nm
Concanavalin A (ConA) - AcalephFluor568	575 nm	598 nm
Concanavalin A (ConA) - AcalephFluor594	593 nm	614 nm
Concanavalin A (ConA) - AcalephFluor633	630 nm	650 nm
Concanavalin A (ConA) - AcalephFluor647	648 nm	664 nm
Concanavalin A (ConA) - AcalephFluor660	663 nm	682 nm
Concanavalin A (ConA) - AcalephFluor680	681 nm	698 nm
Concanavalin A (ConA) - AcalephFluor750	750 nm	777 nm