

# alpha-Hydroxybutyrate Dehydrogenase Microplate Assay Kit User Manual

Catalog # CAK1303

(Version 1.1A)

Detection and Quantification of alpha-Hydroxybutyrate Dehydrogenase ( $\alpha$ -HBDH) Activity in Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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## I. INTRODUCTION

 $\alpha$ -hydroxybutyrate dehydrogenase ( $\alpha$ -HBDH) is an auxiliary marker of myocardial injury, which was reported to have an increased specificity for detecting myocardial injury, starting to increase 8-12 h after damage, reaching peak serum concentrations after 48-72 h and returning to baseline after 7-14 days.

alpha-Hydroxybutyrate Dehydrogenase Microplate Assay Kit provides a simple and direct procedure for measuring alpha-Hydroxybutyrate Dehydrogenase activity in a variety of samples. In this colorimetric alpha-Hydroxybutyrate Dehydrogenase quantification assay, alpha-Hydroxybutyrate Dehydrogenase reduces NAD to NADH, which then interacts with a specific probe to produce a color. The rate of decrease in the absorbency at 450 nm, is a measure of alpha-Hydroxybutyrate Dehydrogenase activity.



# **II. KIT COMPONENTS**

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	10 ml x 1	4 °C
Substrate I	Powder x 1	-20 °C
Substrate II	1 ml x 1	4 °C
Dye Reagent A	Powder x 1	4 °C
Dye Reagent B	1 ml x 1	4 °C
Standard	Powder x 1	4 °C
Positive Control	Powder x 1	-20 °C
Technical Manual	1 Manual	

## Note:

Dye Reagent A: add 9 ml distilled water to dissolve before use, mix, store at 4°C.

Substrate I: add 1 ml Reaction Buffer to dissolve before use, store at -20 °C.

**Standard**: add 1 ml distilled water to dissolve before use; then add 0.15 ml into 0.85

ml distilled water, the concentration will be 300  $\mu$ mol/L, store at -20 °C.

Positive Control: add 0.1 ml distilled water to dissolve before use, store at -80 °C.



# III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 450 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Ice
- 7. Centrifuge
- 8. Timer

## IV. SAMPLE PREPARATION

## 1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

# 2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For serum or plasma samples

Detect directly.



## V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Control	Standard	Blank	Positive		
					Control		
Sample	10 μΙ						
Standard			100 μΙ				
Positive Control					10 μΙ		
Reaction Buffer	70 μΙ	70 μΙ			70 μΙ		
Substrate I	10 μΙ	10 μΙ			10 μΙ		
Substrate II	10 μΙ	10 μΙ			10 μΙ		
Distilled water		10 μΙ		100 μΙ			
Mix.							
Dye Reagent A	90 μΙ	90 μΙ	90 μΙ	90 μΙ	90 μΙ		
Dye Reagent B	10 μΙ	10 μΙ	10 μΙ	10 μΙ	10 μΙ		
Mix, incubate at room temperature for 5 minutes, record absorbance measured at							

## Note:

450 nm.

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.
- 3) Reagents must be added step by step, can not be mixed and added together.



## VI. CALCULATION

**Unit Definition**: One unit of  $\alpha$ -HBDH activity is defined as the enzyme reduce 1  $\mu$ mol NADH per minute.

1. According to the volume of serum or plasma

$$\alpha$$
-HBDH (U/mI) = (C<sub>Standard</sub> × V<sub>Standard</sub>) × (OD<sub>Control</sub> - OD<sub>Sample</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / 
$$V_{Sample} / T$$
= 0.6 × (OD<sub>Control</sub> - OD<sub>Sample</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>)

2. According to the protein concentration of sample

$$\begin{array}{l} \alpha\text{-HBDH (U/mg)} = \left( C_{Standard} \times V_{Standard} \right) \times \left( OD_{Control} - OD_{Sample} \right) / \left( OD_{Standard} - OD_{Blank} \right) / \\ \\ \left( V_{Sample} \times C_{Protein} \right) / T \\ \\ = 0.6 \times \left( OD_{Control} - OD_{Sample} \right) / \left( OD_{Standard} - OD_{Blank} \right) / C_{Protein} \\ \end{array}$$

3. According to the weight of sample

$$\alpha$$
-HBDH (U/g) = (C<sub>Standard</sub> × V<sub>Standard</sub>) × (OD<sub>Control</sub> - OD<sub>Sample</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / (W × V<sub>Sample</sub> / V<sub>Assay</sub>) / T
$$= 0.6 \times (ODControl - ODSample) / (ODStandard - ODBlank) / W$$

4. According to the quantity of cell or bacteria

$$\alpha$$
-HBDH (U/10<sup>4</sup>) = (C<sub>Standard</sub> × V<sub>Standard</sub>) × (OD<sub>Control</sub> - OD<sub>Sample</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / (N   
× V<sub>Sample</sub> / V<sub>Assay</sub>) / T   
= 0.6 × (OD<sub>Control</sub> - OD<sub>Sample</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / N

 $C_{Standard}$ : the concentration of standard, 300  $\mu$ mol/L = 0.3  $\mu$ mol/ml;

C<sub>Protein</sub>: the protein concentration, mg/ml;

W: the weight of sample, g;

V<sub>Sample</sub>: the volume of sample, 0.01 ml;

V<sub>Standard</sub>: the volume of standard, 0.1 ml;

V<sub>Assav</sub>: the volume of Assay buffer, 1 ml;

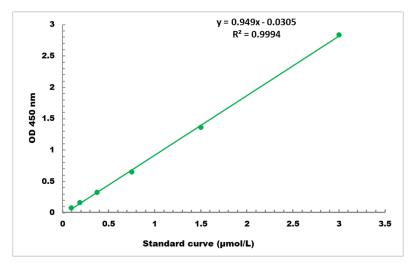
T: the reaction time, 5 minutes;

N: the quantity of cell or bacteria,  $N \times 10^4$ .

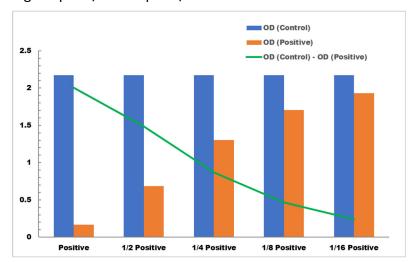


## VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 3 μmol/L - 300 μmol/L



Positive Control reaction in 96-well plate assay with decreasing the concentration

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

# IX. NOTES