



Amylose
Colorimetric Microplate Assay Kit
User Manual

Catalog # CAK1178

(Version 1.4C)

Detection and Quantification of Amylose content in Tissue extracts,
Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.

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I. INTRODUCTION

Amylose is a polysaccharide made of α -D-glucose units, bonded to each other through $\alpha(1\rightarrow4)$ glycosidic bonds. It is one of the two components of starch, making up approximately 20-30%. Amylose is less soluble in water than the other component amylopectin. Because of its tightly packed helical structure, amylose is more resistant to digestion than other starch molecules and is therefore an important form of resistant starch.

AmyloseColorimetric Microplate Assay Kit is a sensitive assay for determining amylose content in various samples. The pure blue is produced according to the action of amylose and iodine reagent. The measurement wavelength and reference wavelength of the amylose were 630nm.

II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay BufferI	30 ml x 4	4 °C
Assay BufferII	30 ml x 4	4 °C
Reaction Buffer A	10 ml x 1	4 °C
Reaction Buffer B	8 ml x 1	4 °C
Dye Reagent	1 ml x 1	4 °C, keep in dark
Standard	Powder x 1	4 °C
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Note:

Standard: add 1 ml Assay Buffer II to dissolve before use, the concentration will be 8 mg/ml.

III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 630nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Centrifuge
6. Timer
7. Mortar

IV. SAMPLE PREPARATION

1. For tissue samples

Weigh out 0.01 g tissue, homogenize with 1 ml Assay Buffer I, then transfer all the lysate to the microtube, centrifuged at 4000g for 10 minutes, discard the supernatant; then add 1 ml Assay Buffer II, warm at 80 °C for 10 mins, centrifuged at 4000g for 10 minutes, take the supernatant into a new centrifuge tube for detection.

V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank
Sample	10 μ l	--	--
Standard	--	10 μ l	--
Distilled water	--	--	10 μ l
Reaction Buffer A	100 μ l	100 μ l	100 μ l
Reaction Buffer B	80 μ l	80 μ l	80 μ l
Dye Reagent	10 μ l	10 μ l	10 μ l
Mix, wait for 5 minutes, record absorbance measured at 630nm.			

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples.
For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.
- 3) Reagents must be added step by step, can not be mixed and added together.

VI. CALCULATION

1. According to the volume of sample

$$\text{Amylose (mg/ml)} = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}})}{(OD_{\text{Standard}} - OD_{\text{Blank}})} \times \frac{V_{\text{Sample}}}{V_{\text{Assay}}}$$

$$= 8 \times (OD_{\text{Sample}} - OD_{\text{Blank}}) / (OD_{\text{Standard}} - OD_{\text{Blank}})$$

2. According to the weight of sample

$$\text{Amylose (mg/g)} = \frac{(C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Blank}})}{(OD_{\text{Standard}} - OD_{\text{Blank}})} \times \frac{V_{\text{Sample}} \times W}{V_{\text{Assay}}}$$

$$= 8 \times (OD_{\text{Sample}} - OD_{\text{Blank}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) \times W$$

C_{Standard} : the standard concentration, 8mg/ml;

W : the weight of sample, g;

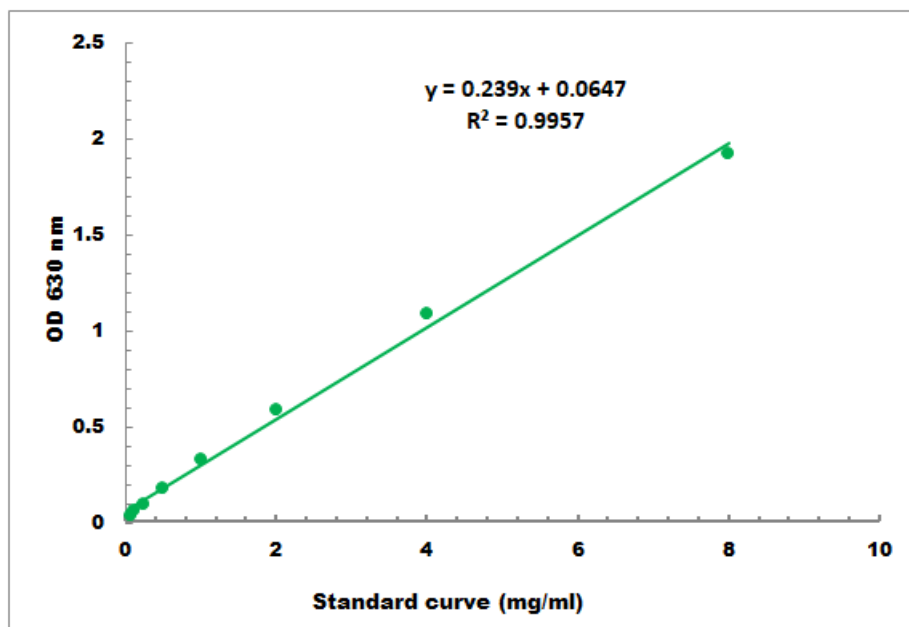
V_{Assay} : the volume of Assay bufferII, 1 ml

V_{Standard} : the volume of standard, 10 μ l;

V_{Sample} : the volume of sample, 10 μ l.

VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.8 mg/ml - 8 mg/ml

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES