

NAD-Malate Dehydrogenase Activity Colorimetric Microplate Assay Kit User Manual

Catalog # CAK1142

(Version 1.4B)

Detection and Quantification of NAD-Malate Dehydrogenase

Activity in Serum, Plasma, Tissue extracts, Cell lysate, Cell culture

media and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



I. INTRODUCTION	2
II. KIT COMPONENTS	3
III. MATERIALS REQUIRED BUT NOT PROVIDED	3
IV. SAMPLE PREPARATION	4
V. ASSAY PROCEDURE	5
VI. CALCULATION	6
VII. TYPICAL DATA	7
VIII. TECHNICAL SUPPORT	7
IX NOTES	7



I. INTRODUCTION

NAD-Malate Dehydrogenase (EC 1.1.1.38) (NAD-MDH) is an enzyme that reversibly catalyzes the oxidation of malate to oxaloacetate using the reduction of NAD+ to NADH. This reaction is part of many metabolic pathways, including the citric acid cycle.

NAD-Malate Dehydrogenase Activity Colorimetric Microplate Assay Kit provides a simple and direct procedure for measuring NAD-malate dehydrogenase activity in a variety of samples. In this kit, NAD-malate dehydrogenase catalyzed reduction of malate in which the formed NADH reduces a formazan reagent. The rate of increase in the absorbency at 450 nm, is a measure of NAD-malate dehydrogenase activity.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	10 ml x 1	4 °C
Substrate	Powder x 1	-20 °C
Dye Reagent A	Powder x 1	4 °C
Dye Reagent B	1 ml x 1	4 °C
Standard	Powder x 1	-20 °C
Positive Control	Powder x 1	-20 °C
Technical Manual	1 Manual	

Note:

Substrate: add 1 ml Reaction Buffer to dissolve before use.

Dye Reagent A: add 9 ml distilled water to dissolve before use, mix, store at 4°C.

Standard: add 1 ml distilled water to dissolve before use; then add 0.2 ml into 0.8 ml distilled water, the concentration will be 400 μ mol/L.

Positive Control: add 1 ml assay buffer to dissolve before use, then add 0.1 ml into 0.8 ml assay buffer, mix. Aliquot & store at -80 °C. Use within one month.



III. MATERIALS REQUIRED BUT NOT PROVIDED

1. N	/licroplate	reader to	read	absorbance	at 450	nm
------	-------------	-----------	------	------------	--------	----

- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Ice

IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For liquid samples

Detect directly.



V. ASSAY PROCEDURE

Warm all regents to room temperature before use.

Add following reagents into the microplate:

Reagent	Sample	Control	Standard	Blank	Positive
					Control
Reaction Buffer	80 μΙ	80 μΙ			80 μΙ
Sample	10 μΙ				
Standard			100 μΙ		
Distilled water		10 μΙ		100 μΙ	
Positive Control					10 μΙ
Substrate	10 μΙ	10 μΙ			10 μΙ
Dye Reagent A	90 μΙ	90 μΙ	90 μΙ	90 μΙ	90 μΙ
Dye Reagent B	10 μΙ	10 μΙ	10 μΙ	10 μΙ	10 μΙ

Mix, keep in dark for 5 minutes at room temperature, record absorbance measured at 450 nm.

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more samples into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.
- 3) Reagents must be added step by step, can not be mixed and added together.



VI. CALCULATION

Unit Definition: One unit of NAD-MDH activity is defined as the enzyme converts one $1 \mu mol$ of NADH per minute.

1. According to the protein concentration of sample

NAD-MDH (U/mg) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) /$$

$$(V_{Sample} \times C_{Protein}) / T$$

$$= 0.8 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / C_{Protein}$$

2. According to the weight of sample

NAD-MDH (U/g) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) /$$

$$(V_{Sample} \times W / V_{Assay}) / T$$

$$= 0.8 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / W$$

3. According to the quantity of cells or bacteria

NAD-MDH (U/10⁴) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} \times N / V_{Assay}) / T$$

= $0.8 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / N$

4. According to the volume of serum or plasma

NAD-MDH (U/mI) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / V_{Sample} / T$$

= $0.8 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank})$

 $C_{Standard}$: the standard concentration, 400 μ mol/L = 0.4 μ mol/ml;

 $V_{Standard}$: the volume of standard, 100 μ l = 0.1 ml;

V_{Sample}: the volume of sample, 0.01 ml;

C_{Protein}: the protein concentration, mg/ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria, $N \times 10^4$;

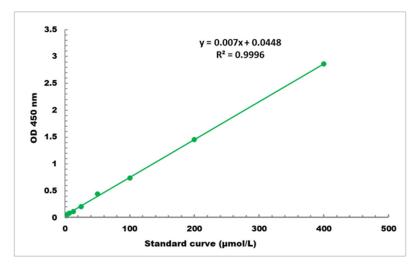
V_{Assay}: the volume of Assay buffer, 1 ml;

T: the reaction time, 5 minutes.

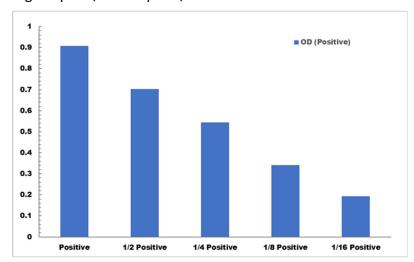


VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 4 μmol/L - 400 μmol/L



Positive Control reaction in 96-well plate assay with decreasing the concentration

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES