



# **Trehalose Microplate Assay Kit**

## **User Manual**

**Catalog # CAK1029**

(Version 2.4H)

Detection and Quantification of Trehalose Content in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

**For research use only. Not for diagnostic or therapeutic procedures.**

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## I. INTRODUCTION

Trehalose is a naturally occurring disaccharide containing two glucose molecules bound in an  $\alpha,\alpha$ -1,1 linkage. This structure results in a chemically stable, non-reducing sugar with many important functional characteristics. Trehalose is commonly found in nature, provides a source of energy, and has been shown to be a primary factor in stabilising organisms during times of freezing, drying and heating. Trehalose Microplate Assay Kit provides a simple and direct procedure for measuring trehalose content in a variety of samples. Trehalose is hydrolysed to glucose by trehalase, and the glucose is oxidized by glucose Oxidase, and can be measured at a colorimetric readout at 505 nm.

## II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer I	10 ml x 1	4 °C
Assay Buffer II	Powder x 1	4 °C
Assay Buffer III	10 ml x 1	4 °C
Enzyme I	30 µl x 1	4 °C
Enzyme II	Powder x 1	-20 °C
Diluent	10 ml x 1	4 °C
Dye Reagent	Powder x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

### Note:

**Assay Buffer II:** add 10 ml Assay Buffer I to dissolve before use.

**Enzyme I:** add 1 ml Diluent to dissolve before use.

**Enzyme II:** add 8 ml Diluent to dissolve before use.

**Dye Reagent:** add 20 ml distilled water to dissolve before use.

**Standard:** add 1 ml distilled water to dissolve before use, then add 0.05 ml into 0.45 ml distilled water, the concentration will be 1 mmol/L.

### **III. MATERIALS REQUIRED BUT NOT PROVIDED**

1. Microplate reader to read absorbance at 505 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer
8. Convection oven

### **IV. SAMPLE PREPARATION**

1. For liquid samples

Add 0.1 ml sample and 0.1 ml Assay Buffer II to the tube, mix on a vortex mixer, keep at 40 °C for 30 minutes; then add 0.1 ml Assay Buffer III to the tube, a vigorous effervescence should be observed, mix on a vortex mixer.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml distilled water, centrifuged at 12000g for 10 minutes. Add 0.1 ml the supernatant and 0.1 ml Assay Buffer II to a new tube, mix on a vortex mixer, keep at 40 °C for 30 minutes; then add 0.1 ml Assay Buffer III to the tube, a vigorous effervescence should be observed, mix on a vortex mixer.

## V. ASSAY PROCEDURE

Add following reagents in the microplate:

Reagent	Sample	Control	Standard	Blank
Sample	50 $\mu$ l	50 $\mu$ l	--	--
Standard	--	--	50 $\mu$ l	--
Distilled water	--	10 $\mu$ l	--	50 $\mu$ l
Enzyme I	10 $\mu$ l	--	10 $\mu$ l	10 $\mu$ l
Enzyme II	40 $\mu$ l	40 $\mu$ l	40 $\mu$ l	40 $\mu$ l
Dye Reagent	100 $\mu$ l	100 $\mu$ l	100 $\mu$ l	100 $\mu$ l
Mix, put the plate into the convection oven, 37 °C for 30 minutes, record absorbance measured at 505 nm.				

### Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples.  
For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.
- 3) Reagents must be added step by step, can not be mixed and added together.

## VI. CALCULATION

### 1. According to the volume of sample

$$\begin{aligned} \text{Trehalose (mmol/L)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Control}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) / \\ &V_{\text{Sample}} \times n \\ &= 3 \times (OD_{\text{Sample}} - OD_{\text{Control}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) \end{aligned}$$

### 2. According to the weight of sample

$$\begin{aligned} \text{Trehalose (mmol/g)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (OD_{\text{Sample}} - OD_{\text{Control}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) / \\ &(W \times V_{\text{Sample}} / V_{\text{Assay}}) \times n \\ &= 3 \times (OD_{\text{Sample}} - OD_{\text{Control}}) / (OD_{\text{Standard}} - OD_{\text{Blank}}) / W \end{aligned}$$

$C_{\text{Standard}}$ : the concentration of standard, 1 mmol/L;

$W$ : the weight of sample, g;

$V_{\text{Assay}}$ : the volume of distilled water, 1 ml;

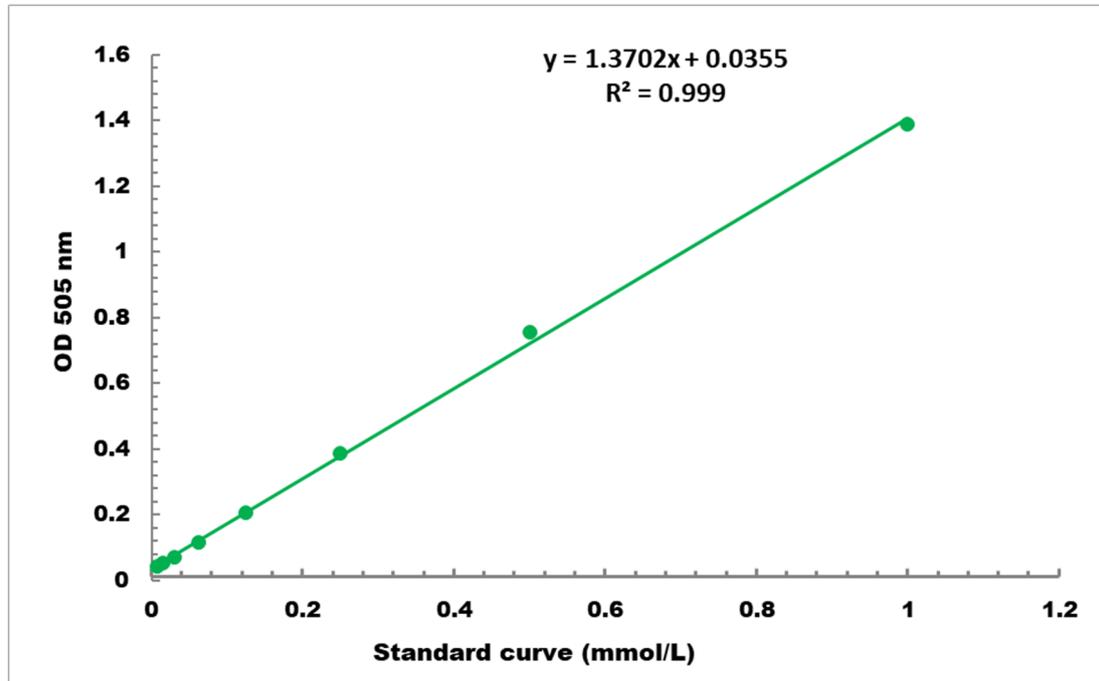
$V_{\text{Standard}}$ : the volume of standard, 0.05 ml;

$V_{\text{Sample}}$ : the volume of sample, 0.05 ml;

$n$ : dilution factor,  $n=3$ .

## VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.01 mmol/L - 1 mmol/L

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to [www.cohesionbio.com](http://www.cohesionbio.com) or contact us at [techsupport@cohesionbio.com](mailto:techsupport@cohesionbio.com)

## IX. NOTES